

Synteny Based Analysis to Proffer a Non-drug Resistant Target Against *Porphyromonas gingivalis*

Ashok Kumar¹, Charu Sood¹, Aditi Rathee², and Amit Pal^{*3}

¹Centre for System Biology and Bioinformatics, Panjab University, Chandigarh, India

²Department of Microbiology, Panjab University, Chandigarh, India

³Department of Biochemistry, PGIMER, Chandigarh, India

*Corresponding author: Amit Pal, Department of Biochemistry, P.G.I.M.E.R., Chandigarh, India 160012, Tel: +91-8427007109, E-mail: maximus1134@gmail.com, apal1134@gmail.com

Citation: Ashok Kumar, Charu Sood, Aditi Rathee, Amit Pal (2015) Synteny Based Analysis to Proffer a Non-drug Resistant Target Against *Porphyromonas gingivalis*. J Comp Sys Bio 1(1): 105. doi: 10.15744/2455-7625.1.105

Received Date: August 01, 2015 **Accepted Date:** August 29, 2015 **Published Date:** August 31, 2015

Abstract

Porphyromonas gingivalis is an anerobic, gram negative, bacterium which is a natural inhabitant of oral microbiome. Under unhealthy oral hygiene conditions the bacterium becomes highly destructive (photobiont) and leads to periodontitis (inflammation of periodontium). Several drugs like chlorhexidine, monocycline effectively reduce the viability of biofilms formed by *Porphyromonas gingivalis* but, it is impossible to destroy biofilms completely. As the bacterium evolves continuously it develops drug resistance due to events like horizontal gene transfer and its biofilm mode of living. Therefore no drug is very efficient in treating Periodontitis. We have done an *In silico* Synteny based comparative analysis to find a druggable target which remains unchanged in various *Porphyromonas gingivalis* strains. Dipeptidylaminopeptidase has been identified one such potential target which is responsible for virulence in *Porphyromonas gingivalis*, and is functionally and structurally conserved. Therefore, according to best knowledge of authors a drug developed against this target will be effective in treating periodontitis caused by all evolved forms of bacterium.

Keywords: *Porphyromonas gingivalis*; Periodontitis; Drug resistance; Synteny.

Abbreviations: DPP IV: Dipeptidylaminopeptidase IV; BLAST: Basic Local Alignment Search Tool; NCBI : National Center for Biotechnology Information; RMSD: Root Mean Square Deviation; ATCC: American Type Culture Collection; MSA: Multiple Sequence alignment

Introduction

Periodontitis is inflammatory diseases caused by peridontopathic bacteria, majorly acquires gingival crevice, often causing gums to shrink and loosening of teeth. It is apparently caused by specific gram negative anaerobic bacteria [1], *Porphyromonas gingivalis*, *Treponema denticola*, and Tannerella (formerly Bacteroides) forsythia more commonly termed as the “red complex”[2,3]. *Porphyromonas gingivalis* is a natural member of more than 500 bacterial species that live in oral microbiome, yet it becomes immensely catastrophic (termed pathobiont) and proliferates to high cell numbers in periodontal lesions: this is credited to its arsenal of specialized virulence factors. Gingival epithelial cells, the outer lining of the gingival mucosa, which function as an innate part of the innate immune system, are among the first host cells colonized by *Porphyromonas gingivalis* [4]. *Treponema denticola* leads to necrotizing ulcerative gingivitis after getting adhered to fibroblasts, epithelial cells, as well as to extracellular matrix components present in periodontal tissues. It possesses outer-sheath-associated peptidases, chymotrypsin-like and trypsin-like proteinases, hemolytic and hemagglutinating activities, adhesins as virulence factors which aid in early periodontitis [5]. *T.forsythia* is another anaerobic, Gram-negative bacterium isolated from the gingival sulci and periodontal pockets of patients with periodontitis [3]. Several *T.forsythia* virulence factors have been identified [6], including, trypsin-like [7] and PrtH proteases [8], sialidases SiaH [9] and NanH [10], a leucine-rich repeat cell-surface-associated and secreted protein BspA [11], an apoptosis-inducing activity [12], alpha-D-glucosidase and N-acetyl-beta-glucosaminidase [13], a hemagglutinin [14], components of the bacterial S-layer [15].

Within group of red complex, *P. gingivalis* and *T. denticola* are frequently found together in diseased sites [3], but *T. forsythia*, may not co localize spatially with them [16,17]. Also *T. denticola* do not form homotypic biofilms. It is *P. gingivalis* which acted as an initial colonizer of the substratum, enabling subsequent incorporation of *T. denticola* [18,19]. Gingipains (*P. gingivalis*) and dentilisin (*T. denticola*) have happened to be involved in the co aggregation [19,20]. Due to the fastidious growth requirement for culturing *T. forsythia* and the genetic manipulations of this organism are difficult to perform, this organisms is understudied [21]. Among the species of red complex, *P. gingivalis* has happened to be a major pathogen, with well-defined virulence factors

including cell surface located proteolytic enzymes and adhesins (gingipains) [22,23]. Recent studies have focused on the pathogenic traits of the *Porphyromonas gingivalis*. A pioneering work by scientist suggests that *Porphyromonas gingivalis* produces an array of potential virulence factors viz capsule, outer membrane protein and gingipains etc. It possesses strong proteolytic activity, which is considered to be associated with tissue destruction. It cleaves epithelial junction proteins, which suggests that it invades deep into tissue via paracellular pathway and thereby destroying the connective tissue. Identified proteases are Arg-specific and Lys-specific cysteine proteases, DPPIV [24], prolyltri-peptidyl peptidase (PtpA), and others [25].

There are several ways of treating periodontitis, non-surgical methods like (scaling and root planning), surgical methods (flap surgery, bone and tissue grafts) medication is used with non-surgical treatment but, it has been seen that *Porphyromonas gingivalis* biofilms persist even after treatment with chlorhexidine (chlorinated phenolic disinfectant used commonly in mouthwash for its action against bacteria) [26] and other known antibiotics also. The bacterium continuously evolves since the biofilm it forms with other bacteria provides an ideal niche for the exchange of extrachromosomal DNA which encodes for resistance to antimicrobial agents and therefore making it difficult to treat bacteria [27]. It is known that certain residues that perform key functional and structural roles tend to be preserved by natural selection. In this regard discovering genome level similarity based on modern approach like Synteny Based Comparative Genomics is serving us to find precisely the essential genes which are conserved and play a role in intensification of bacterial infection as well as in promoting drug resistance in case of periodontitis and therefore, can be potential drug targets.

Material and Methods

Selection of genomes and a reference organism

NCBI Genome was used for genome selection of *Porphyromonas gingivalis*. Only the genomes that were completely sequenced (*Porphyromonas gingivalis* ATCC 33277 and *Porphyromonas gingivalis* TDC60) were selected for carrying comparative genome analysis. According to literature and database availability of complete genome information, we opted for *Porphyromonas gingivalis* W83 as a reference genome.

Synteny analysis between *Porphyromonas gingivalis* W83 and its two completely sequenced strains

Microscope server (www.genoscope.cns.fr/agc/microscope/home) [28] was used to carry out the synteny analysis between the three fully sequenced genomes of *Porphyromonas gingivalis*. Firstly the reference genome was selected and loaded into the browser and then the other two strains (*Porphyromonas gingivalis* ATCC 33277 and *Porphyromonas gingivalis* TDC60) were selected. Homologous synteny blocks were identified based on Microscope search. The list of the conserved synteny blocks was downloaded in the form of MS-Excel data.

Prognosis of putative target in homologous non –variable genes

From the synteny data obtained, variable and non- variable genes was filtered based on the observation that a particular gene from the comparable genomes corresponded to only a single gene in the reference genome also, such genes were taken under non- variable genes and the conserved blocks that corresponded to more than one gene in the compared genomes were kept under variable genes. Then, a literature study was performed using PubMed to determine the major virulent factors present in *Porphyromonas gingivalis*. These genes was then searched in non-variable genes and a list of virulent factors were made and each of which was individually studied to find a factor or a potential target that was conserved in all the strains and had role in increasing the pathogenic potential of *Porphyromonas gingivalis*.

Insilico analysis of the putative target

Insilico characterization of target was done by retrieving the nucleotide and amino acid sequences from NCBI. MSA using MUSCLE [29] was done for protein as well as nucleotide sequences.

Validation of the target

The selected target was validated performing a database search using BLAST [30] to see whether the target was specific to the bacterium or it was homologous to some human protein. After BLAST results, conserved domains were predicted using InterPro [31] to see structurally and functionally similar domains in the target sequences from three different strains of *Porphyromonas gingivalis*.

Results

Synteny analysis of selected genomes

Only the completely sequenced genomes of *Porphyromonas gingivalis* were selected. *Porphyromonas gingivalis* W83 was taken as a reference genome and was compared with the other two completely sequenced genomes of *Porphyromonas gingivalis* ATCC 33277, and *Porphyromonas gingivalis* TDC60. Results from Microscope server showed conserved synteny blocks between the reference and other two strains. Colored boxes in the results represent synteny conservations in at least one organism of the related taxon (Figure 1).

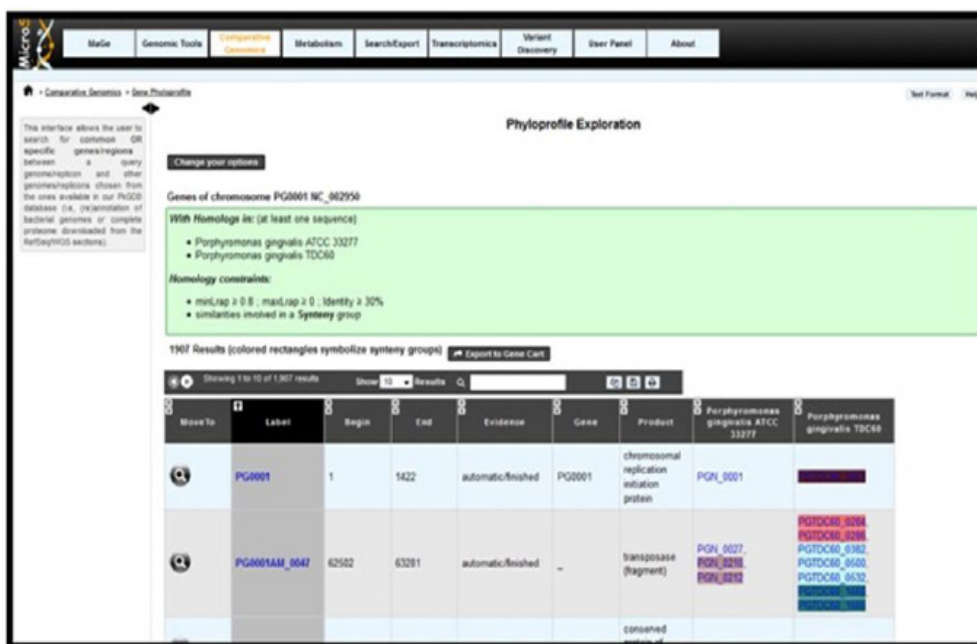


Figure 1: Microscope Synteny homepage

Selection of a putative target from non-variable genes

Literature studies suggested that peptidases in *Porphyromonas gingivalis* were the major reason for its virulence and therefore from the list of non-variable genes (Table 1), another list of all the peptidases was made and analyzed further and out of which DPP IV was selected as a potential target (Table 2).

Begin	End	<i>Porphyromonas gingivalis</i> W83	Product	<i>Porphyromonas gingivalis</i> ATCC 33277	<i>Porphyromonas gingivalis</i> TDC60
1	1422	PG0001	Chromosomal replication initiation protein	PGN_0001	PGTDC60_0001
1435	2007	PG0002	Hexapeptidetransferase	PGN_0002	PGTDC60_0002
2009	3028	PG0003	Hypothetical protein	PGN_0003	PGTDC60_0003
3108	3812	PG0004	NAD-dependent deacetylase	PGN_0004	PGTDC60_0004
3887	5041	PG0005	Hypothetical protein	PGN_0005	PGTDC60_0005
5094	6422	PG0006	Mate efflux family protein	PGN_0006	PGTDC60_0006
11634	14645	PG0011	glycosyl hydrolase	PGN_0009	PGTDC60_0011
11693	15721	PG0012	L-threonine O-3 phosphate decarboxylase	PGN_0010	PGTDC60_0012
15731	16501	PG0013	Hypothetical protein	PGN_0011	PGTDC60_0013

A list of non variables genes was obtained.

Table 1: A set of non -variable genes in *Porphyromonas gingivalis* strains

S.no	Peptidases in <i>Porphyromonas gingivalis</i>	Corresponding genes ids
1.	Signal peptidase like protein	PG2000, PGN_1945, PGTDC60_0273
2.	Signal Peptidase 1	PG2001 , PGN_1946, PGTDC60_0275
3.	M16 family peptidase	PG0088, PGN_1946, PGTDC60_0367
4.	Aminoacyl-histidinedipeptidase	PG0137, PGN_0250, PGTDC60_0414
5.	EndopeptidasePepO	PG0159, PGN_0271, PGTDC60_0510
6.	Zinc carboxypeptidase	PG0232, PGN_0335, PGTDC60_0510
7.	M49 family peptidase	PG0317, PGN_1645, PGTDC60_1437
8.	Dipeptidylaminopeptidase IV	PG0503, PGN_1469, PGTDC60_1620
9.	Peptidase T	PG0445, PGN_1517, PGTDC60_1565
10.	Prolyl oligopeptidase	PG0724, PGN_1776, PGTDC60_18845
11.	Peptidyl-dipeptidaseDcp	PG1789, PGN_0756, PGTDC60_0105
12.	M24 family peptidase	PG0889, PGN_1050, PGTDC60_0186
13.	Cysteine peptidase	PG1788, PGN_1777, PGTDC60_0106

From the list of non- variable genes after literature study peptidases were separated into a different list

Table 2: A list of all peptidases separated from non-variable genes

Target analysis

Nucleotide sequences of the three DPP IV genes from three different strains were analyzed by performing MSA by MUSCLE. The percent identity matrix has identity of 61% between the sequences and hence was found to be homologous to each other. Percent identity matrix of amino acid sequences revealed identity of more than 30% hence was homologous. Amino acid sequences retrieved from NCBI had shown that the amino acid number varied from first sequenced to recently sequenced strains *Porphyromonas gingivalis* W83 had 732 amino acids [32], *Porphyromonas gingivalis* ATCC 33277 had 723 amino acids [33] and *Porphyromonas gingivalis* TDC60 had 709 amino acids [34], respectively. This variation in amino acid numbers could be due to evolution of bacterium and would have led to some loss or gain in function.

Validation of DPP IV as a potential target

In order to make sure that *Porphyromonas gingivalis* DPP IV is specific and is not found in humans, BLAST database search was conducted against non-redundant protein database and results gave no significant similarity with any human protein for three DPP IV sequences. Therefore we conclude that the target selected was specific to *Porphyromonas gingivalis*.

Structural analysis of DPP IV

InterPro gave the two common conserved domains for three DPP IV sequences. IPR029058 (a region in DPP IV predicted as Alpha/Beta hydrolase fold; this structure is common to a number of hydrolytic enzymes of widely differing phylogenetic origin) and IPR002469 [a region in DPP IV as belonging to serine peptidase family (S9)]. This led us to believe that even though the sequence is changing, functionally and structurally important regions are still conserved. Therefore, a drug made against this target will remain effective for all *Porphyromonas gingivalis* strains and since DPP IV is highly conserved it will not show resistance to the drug.

Structure prediction of DPP IV

Secondary structures was predicted using PHD [35] showed that number of alpha helices was increasing in newly sequenced bacteria hence making DPPIV a more stable structure (Table 3). Tertiary structures were generated using SWISS MODEL and were then visualized in Chimera, all the three DPP IV structures predicted were found to be homodimers.

Strains	Alpha helices	Extended strands	Random Coils
<i>Porphyromonas gingivalis</i> W83	14%	33.33%	52.19%
<i>Porphyromonas gingivalis</i> ATCC 33277	25.73%	28.77%	45.50%
<i>Porphyromonas gingivalis</i> TDC 60	26.52%	29.66%	44.43%

Increase in the number of alpha helices and decrease in the number of, extended strands and random coils are seen.

Table 3: PHD results showing variation in terms of secondary structures

Structure validation of DPP IV

Verify3D [36] validated structures generated by SWISS MODEL, all structures obtained profile scored greater than 0.2. The structure was also validated by generating Ramachandran plots of the three models and by calculating RMSD values [37] for the superimposed structures in chimera (Table 4). Maximum residues in Ramachandran plot lied in allowed regions and RMSDs obtained using both Smith Waterman and Needleman Wunch algorithm were less than 2 angstrom.

Algorithm used	RMSD Value W83 with ATCC 33277	RMSD Value of W83 with TDC60
Needleman Wunch Algorithm	1.221	1.238
Smith- Waterman Algorithm	1.221	1.238

RMSD values obtained were less than 0.2 angstroms

Table 4: RMSD values as calculated in Chimera

Discussion

The current study illustrates an emerging field of comparative genomics i.e. synteny based analysis which allows us unmask the conserved gene content and gene order. Identifying Synteny blocks and aligning whole genome both rely on markers across the genomes long enough to make their conservation statistically significant [38]. Till date many conventional therapies have been used in treating periodontitis but eventually fail due its complex multi-factorial etiology [39]. Therapies include mechanical removal of bacterial deposits and calculus from subgingival environment or performing rooting and scaling followed by use of antibacterial agents mainly chlorhexidine, tetracycline, doxycycline and minocycline but local delivery is cost effective. They bind to tooth and reduce bacterial count and dental plaque, out of which chlorhexidine is a gold standard in dentistry [40,41]. Since *P.gingivalis* colonizes first to the tooth surface followed by synergy association with *T. denticola* in establishing biofilms and with *T.forsythia*, it is assumed that if a target is chosen from *P.gingivalis* it will strongly effect the association which the organisms have with other two anaerobes. According to the literature, many protease targets have been reported which accounts for strong proteolytic activity due to which *P. gingivalis* invade into deeper tissues [4]. With the aid of synteny based study, we are able to

deduce one such potential druggable target i.e. DPP1V protease which is involved majorly in pathogenesis of bacterium. This target is species specific and structurally and functionally conserved in all the known three strains of *P. gingivalis*. To wrap up if drug is made against this target of *P. gingivalis* which happened to be primary colonizer, it would staunch enough treat to periodontitis effectively.

Conclusion

Synteny based analysis of *P. gingivalis* has led to testimony of diverse proteases that can be used for potential target for drug design. As various proteases have been noted to play a substantial role in pathogenesis of periodontitis, a systematic approach to develop drug will be propitious for treatment. In this study DPP1V has found to play a decisive role in breaching deeper tissues and biofilm formation which happens to be an imperative factor in periodontitis. It is expected that if drug is developed against this will be roseate in cure.

Acknowledgement

Authors are thankful to Vice Chancellor of Panjab University, Chandigarh, and Coordinator of Centre for Systems biology and Bioinformatics for providing all the facilities to carry out the present work. We acknowledge the free softwares, tools and web servers used in this present study (NCBI genome, Microscope server, MUSCLE, BLAST, InterPro, SWISS model, PHD, Chimera).

References

- Affajee AD, Cugini MA, Dibart S, Smith C, Kent Jr, et al. (1997) Clinical and microbiological features of subjects with adult periodontitis who responded poorly to scaling and root planing. *J Clin Periodontol* 24: 767–76.
- Kasuga Y, Ishihara K, Okuda K (2000) Significance of detection of Porphyromonas gingivalis, Bacteroides forsythus and Treponema denticola in periodontal pockets. *Bull Tokyo Dent Coll* 41: 109–17.
- Socransky SS, Haffajee AD, Cugini MA, Smith C, Kent Jr RL (1998) Microbial complexes in subgingival plaque. *J Clin Periodontol* 25: 134–44.
- Mysak J, Podzimek S, Sommerovav P, Lyuya-Mi Y, Bbartova J, et al. (2014) Porphyromonas gingivalis: Major Periodontopathic Pathogen Overview. *J Immunol Res* 476068.
- Sela MN (2001) Role of Treponema denticola in periodontal diseases. *Critical Review Oral Biol Med* 12: 399–413.
- Sharma A (2010) Virulence mechanisms of Tannerella forsythia. *Periodontolo* 54: 106–16
- Grenier D (1995) Characterization of the trypsin-like activity of Bacteroides forsythus. *Microbiol* 141: 921–6.
- Saito T, Ishihara K, Kato T, Okuda K (1997) Cloning, expression, and sequencing of a protease gene from Bacteroides forsythus ATCC 43037 in Escherichia coli. *Infect Immun* 65: 4888–91.
- Ishikura H, Arakawa S, Nakajima T, Tsuchida N, Ishikawa I (2003) Cloning of the Tannerella forsythensis (Bacteroides forsythus) siaHI gene and purification of the sialidase enzyme. *J Med Microbiol* 52: 1101–7.
- Thompson H, Homer KA, Rao S, Booth V, Hosie AH (2009) An orthologue of Bacteroides fragilis NanH is the principal sialidase in Tannerella forsythia. *J Bacteriol* 191: 3623–8.
- Sharma A, Sojar HT, Glurich I, Honma K, Kuramitsu HK, et al. (1998) Cloning, expression, and sequencing of a cell surface antigen containing a leucine-rich repeat motif from Bacteroides forsythus ATCC 43037. *Infect Immun* 66: 5703–10.
- Nakajima T, Tomi N, Fukuyo Y, Ishikura H, Ohno Y, et al. (2006) Isolation and identification of a cytopathic activity in Tannerella forsythia. *Biochem Biophys Res Commun* 133–9.
- Hughes CV, Malki G, Loo CY, Tanner AC, Ganeshkumar N (2003) Cloning and expression of alpha-D-glucosidase and N-acetyl-beta-glucosaminidase from the periodontal pathogen, Tannerella forsythensis (Bacteroides forsythus). *Oral Microbiol Immunol* 18: 309–12.
- Murakami Y, Higuchi N, Nakamura H, Yoshimura F, Oppenheim FG (2002) Bacteroides forsythus hemagglutinin is inhibited by N-acetylneuraminylactose. *Oral Microbiol Immunol* 17: 125–8.
- Sakakibara J, Nagano K, Murakami Y, Higuchi N, Nakamura H, et al. (2007) Loss of adherence ability to human gingival epithelial cells in S-layer protein-deficient mutants of Tannerella forsythensis. *Microbiol* 153: 866–76.
- Zijngje V, van Leeuwen MB, Degener JE, Abbas F, Thurnheer T, et al. (2010) Oral biofilm architecture on natural teeth. *PLoS One* 5: e9321.
- Wood SR, Kirkham J, Marsh PD, Shore RC, Nattress B, et al. (2000) Architecture of intact natural human plaque biofilms studied by confocal laser scanning microscopy. *J Dent Res* 79: 21–7.
- Yamada M, Ikegami A, Kuramitsu HK (2005) Synergistic biofilm formation by Treponema denticola and Porphyromonas gingivalis. *FEMS Microbiology Letters* 250: 271–7.
- Cogoni V, Morgan-Smith A, Fenno JC, Jenkinson HF, Dymock D (2012) Treponema denticola chymotrypsin-like proteinase (CTLP) integrates spirochaetes within oral microbial communities. *Microbiol* 158: 759–70.
- Ito R, Ishihara K, Shoji M, Nakayama K, Okuda K (2010) Hemagglutinin/ Adhesin domains of Porphyromonas gingivalis play key roles in coaggregation with Treponema denticola. *FEMS Immunol Med Microbio* 60: 251–60.
- Sakakibara J, Nagano K, Murakami Y, Higuchi N, Nakamura H, et al. (2007) Loss of adherence ability to human gingival epithelial cells in S-layer protein-deficient mutants of Tannerella forsythensis. *Microbiol* 153: 866–76.
- Potempa J, Sroka A, Imamura T, Travis J (2003) Gingipains, the major cysteine proteinases and virulence factors of Porphyromonas gingivalis: structure, function and assembly of multidomain protein complexes. *Curr Protein Pept Sci* 4: 397–407.
- Amano A (2003) Molecular interaction of Porphyromonas gingivalis with host cells: implication for the microbial pathogenesis of periodontal disease. *J Periodontol* 74: 90–6.
- Kumagai Y, Yagishita H, Yajima A, Okamoto T, Konishi K (2005) Molecular mechanism for connective tissue destruction by Dipeptidylaminopeptidase IV produced by the periodontal pathogen Porphyromonas gingivalis. *Infect Immun* 73: 2655–64.
- Yilmaz O (2008) The chronicles of Porphyromonas gingivalis: the microbium, the human oral epithelium and their interplay. *Microbiology* 154: 2897–903.

26. Yamaguchi M, Noiri Y, Kuboniwa M, Yamamoto R, Asahi Y, et al. (2013) Porphyromonas gingivalis biofilms persist after chlorhexidine treatment. *Eu J Oral Sci* 121: 162-8.
27. Tribble GD, Kerr JE, Wang BY (2013) Genetic diversity in the oral pathogens Porphyromonas gingivalis: molecular mechanisms and biological consequences. *Future Microbiol* 8: 607-20.
28. Vallenet D, Belda E, Calteau A, Cruveiller S, Engelen S, et al. (2013) MicroScope--an integrated microbial resource for the curation and comparative analysis of genomic and metabolic data. *Nucleic Acids Res* 41: D636-47.
29. Edgar RC (2004) MUSCLE: Multiple sequence alignment with high accuracy and high throughput. *Nucleic Acids Res* 32: 1792-7.
30. Altschul SE, Gish W, Miller W, Myers EW, Lipman DJ (1990) Basic local alignment search tool. *J Mol Biol* 215: 403-10.
31. Hunter S, Apweiler R, Attwood TK, Bairoch A, Bateman A, et al. (2009) The integrative protein signature database. *Nucleic Acids Research* 37: D211-5.
32. Nelson KE, Fleischmann RD, DeBoy RT, Paulsen RT, Fouts DE, et al. (2003) Complete genome sequence of bacterium of oral pathogenic bacterium. Porphyromonas gingivalis strain W83. *J bacterial* 18: 5591-601.
33. Naritto M, Hirakawa H, Yamashita A, Ohara N, Shoji M, et al. (2008) Determination of genomesquence of Porphyromonas gingivalis Strain ATCC 33277 and Genomic Comparison with Strain W83 revealed Extensive Genome Rearrangements in P.gingivalis. *DNA Res* 15: 215-25.
34. Watanabe T, Maruyama F, Nozawa T, Aoki A, Okano S, et al. (2011) Complete genome sequence of bacterium Porphyromonas gingivalis TDC60, which cause periodontal disease. *J Bacteriol* 16: 4259-60.
35. Rost B, Sander C, Schneider R (1994) PHD--an automatic mail server for protein secondary structure prediction. *Comput Appl Biosci* 10: 53-60.
36. Eisenberg D, Lüthy R, Bowie JU (1997) VERIFY3D: assessment of protein models with three-dimensional profiles. *Methods Enzymol* 277: 396-404.
37. Maiorov VN, Crippen GM (1994) Significance of root-mean-square deviation in comparing three-dimensional structures of globular proteins. *J Mol Biol* 235: 625-34.
38. Ghiurcuta CG, Moret BM (2014) Evaluating synteny for improved comparative studies. *Bioinformatics* 30: i9-18.
39. Prakasam A, Elavarasu SS, Natarajan RK (2012) Antibiotics in the management of aggressive periodontitis. *J Pharm Bioallied Sci* 4: S252-5.
40. Luciana M Shaddox, Clay B Walker (2010) Treating chronic periodontitis: current status, challenges, and future directions. *Clinical, Cosmetic and Investigational Dentistry* 2: 79-91.
41. Vandekerckhove BN, Quirynen M, van Steenberghe D (1997) The use of tetracycline-containing controlled-release fibers in the treatment of refractory periodontitis. *J Periodontol* 68: 353-61.

Submit your next manuscript to Annex Publishers and benefit from:

- ▶ Easy online submission process
- ▶ Rapid peer review process
- ▶ Online article availability soon after acceptance for Publication
- ▶ Open access: articles available free online
- ▶ More accessibility of the articles to the readers/researchers within the field
- ▶ Better discount on subsequent article submission

Submit your manuscript at

<http://www.annepublishers.com/paper-submission.php>